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ORIGINAL ARTICLE

Synthesis of new derivatized pyrazole based ligands and their catecholase activity studies

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Abstract A synthesis of three new tripodal ligands: 3-[bis-(3,5-dimethyl-pyrazol-1-ylmethyl)-amino]-propan-1-ol **L1**, 3-[bis-(5-methyl-3-carbomethoxy-pyrazol-1-ylmethyl)-amino]-propan-1-ol, **L2** and 3-[bis-(5-methyl-3-carboethoxy-pyrazol-1-ylmethyl)-amino]-propan-1-ol **L3** is reported. The *in situ*-generated copper(II) complexes of three new compounds (**L1**–**L3**) were examined for their catalytic activities and were found to catalyse the oxidation reaction of catechol to *o*-quinone with the atmospheric dioxygen. These activities depend on the nature of the ligand and the copper salts.

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1. Introduction

It is now well-documented that copper containing metallo-proteins play a very important role in transport, activation, and metabolism of dioxygen in living organisms (Albada et al., 2007; Chen and Solomon, 2004; Decker and Tuzek, 2000; Holm et al., 1996). A notable advance in the understanding of the properties of these proteins has been achieved through the comparison of synthetic models to the naturally occurring molecules (Decker et al., 2000; Kitajima and Moro-oka, 1994; Van Gelder et al., 1997). Several catechol derivative substrates were used in the literature to understand the mechanisms of oxidase enzyme research (Gerdmann et al., 2002; Rompel et al., 1999). It was observed that the catalytic activities of the complexes are not only dependent on the organic ligand but also on the type of inorganic anion coordinated to the copper center (Koval et al., 2006). In this paper, we report the synthesis of three new pyrazolyl ligands. The copper(II) *in situ*-generated complexes of these new products, were examined as catalysts to-

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ward atmospheric dioxygen oxidation reaction of catechol to *o*-quinone.

2. Results and discussion

2.1. Synthesis

Pyrazolyl and triazolyl derivatives N–C–N junction ligands **L1–L3** (Scheme 1) were prepared using two different methods. The first one consists of condensation of 1-(hydroxymethyl)-3,5-dimethylpyrazole (Dvoretzky and Richter, 1950), 1-hydroxymethyl-5-methyl-1H-pyrazole-3-carboxylic acid methyl ester (Touzani et al., 2000), 1-hydroxymethyl-5-methyl-1H-pyrazole-3-carboxylic acid ethyl ester (Touzani et al., 1999) with a series of alcohol amines such as aminopropanol in 2:1 ratio at room temperature during four days in acetonitrile (Touzani et al., 2001). In the second method, the similar reactions were carried out with some drops of solvent at 60 °C for 4 h. The target products were isolated with good yields in the two methods from 75% to 89% (Table 1).

All the new compounds were characterized by ¹H NMR, ¹³C NMR and mass spectrometry. The proton NMR spectra of **L1** product, show two signals at 4.90 and 4.36 ppm corresponding to methylene protons (N–CH₂–N). When the pyrazolic rings are linked to the ester (–CO₂R) moieties (**L2**, **L3**), these proton signals appears between 5.06 and 5.29 ppm.

2.2. Catecholase studies

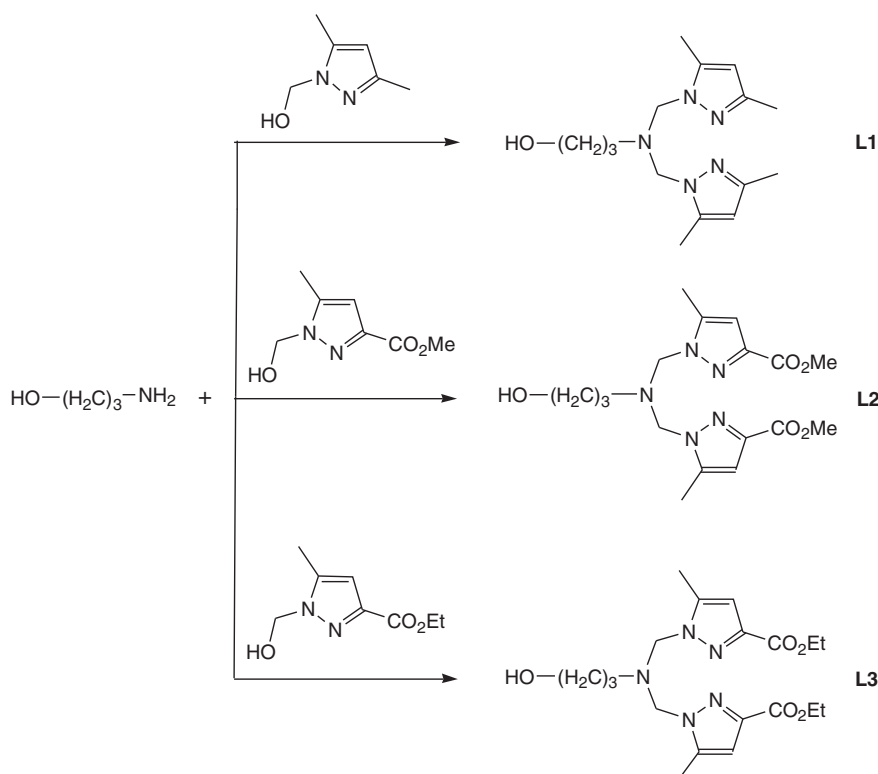
The progress of the catechol oxidation reaction is conveniently followed monitoring the strong absorbance peak of quinone in the UV/Vis spectrophotometer (Scheme 2).

The metal complex solution and catechol reductant were added together in the spectrophotometric cell at 25 °C (El Kodadi et al., 2008; Bouabdallah et al., 2007; Boussalah et al., 2009a,b). Formation of *o*-quinone was monitored by the increase in absorbance at 390 nm as a function of time. Figs. 1–6 show the absorbance versus time spectrum for the first 60 min of the oxidation reaction, while the rates are shown in Table 2. In all cases, catecholase activity was noted.

As can be seen from Table 2, all of the complexes with pyrazolyl ligands catalyze the oxidation reaction of catechol to *o*-quinone with the rate varying from a high of 0.0289 μmol substrate per mg catalyst per min for the **L1**[CuSO₄] complex to a low of 0.0018 μmol substrate per mg catalyst per min for **L1**[CuCl₂]. These rates are comparable to the values reported by (Malachowski et al., 1996) from 0.018 to 0.186 μmol substrate per mg catalyst per minute, for the similar tripodal ligands. The catalytic activity depends strongly on both the R substituent and the type of inorganic anion. The copper complexes of ligand **L1** were observed to be the lowest active, except in the case when we have used SO₄^{2–} anion. The order of reactivity for the oxidation of catechol by CuCl₂ and Cu(NO₃)₂ complexes is **L2** > **L3** > **L1**. The order of reactivity for the oxidation of catechol by Cu(CH₃COO)₂ complexes is **L1** with the other ligands the rate of activity decreases strongly. The order of reactivity for the oxidation of catechol by CuSO₄ complexes is **L1** > **L2** > **L3**.

3. Conclusion

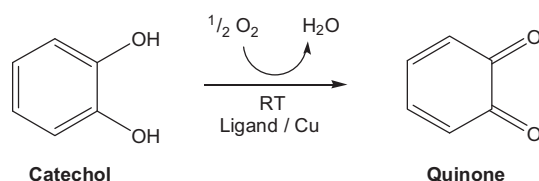
We report the synthesis of amino acid functional tridentate ligands with good and excellent yields. The oxidation of catechol is very efficient to give quinone by complexes of



Scheme 1

Table 1 Comparison between reaction yields.

Products	Method 1 (%)	Method 2 (%)
L1	80	87
L2	78	88
L3	75	89

**Scheme 2**

copper(II) with four functional tripodal pyrazole ligands. The complexes of copper(II) were generated *in situ*. We have demonstrated that the nature of the ester side chain has a large effect on the oxidation reaction rate. The study of various copper(II) salts shows that the catalytic activities are most controlled by the nature of the anion, too.

4. Experimental section

4.1. Apparatus

NMR spectra (^1H , ^{13}C) were recorded on a BRUKER 300 (operating at 300.13 MHz for ^1H , 75.47 MHz for ^{13}C) spectrometer. Chemical shifts are listed in ppm and are reported relative to tetramethylsilane (^1H , ^{13}C). Residual solvent peaks being used as internal standard. The mass spectra have been obtained on a Micromass LCT spectrometer.

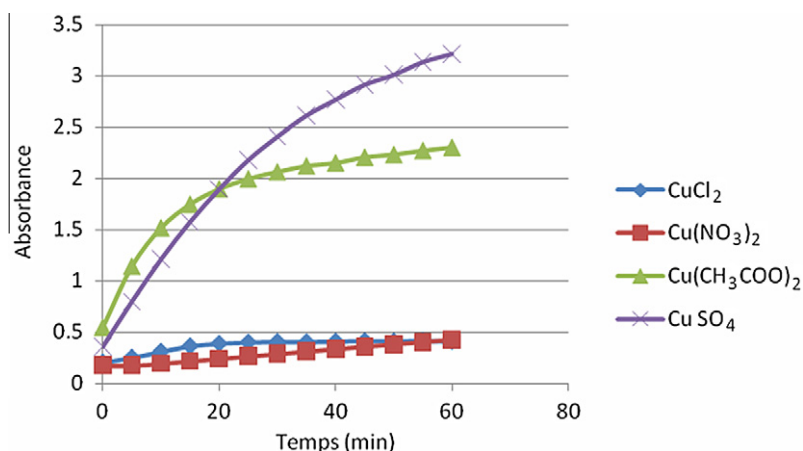
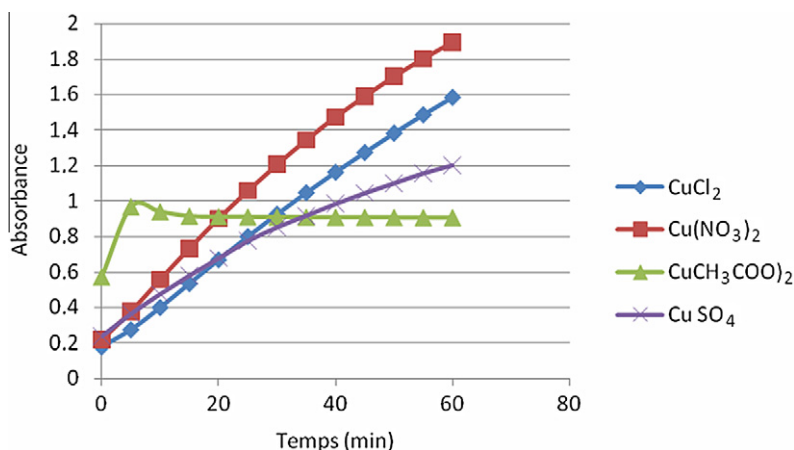
4.2. General procedure

4.2.1. Method 1

A mixture of 1-(hydroxymethyl)-3,5-dimethylpyrazole; 1-hydroxymethyl-5-methyl-1H-pyrazole-3-carboxylic acid methyl ester and 1-hydroxymethyl-5-methyl-1H-pyrazole-3-carboxylic acid ethyl ester with an alcohol amine in 2:1 ratio was stirred for four days in acetonitrile.

4.2.2. Method 2

A mixture of one equivalent of the appropriate amines (10 mmol) and two equivalents (20 mmol) of hydroxymethyl-derivatives in acetonitrile (1 mL), was heated in a water bath

**Figure 1** Oxidation of catechol by complexes of ligand **L1**.**Figure 2** Oxidation of catechol by complexes of ligand **L2**.

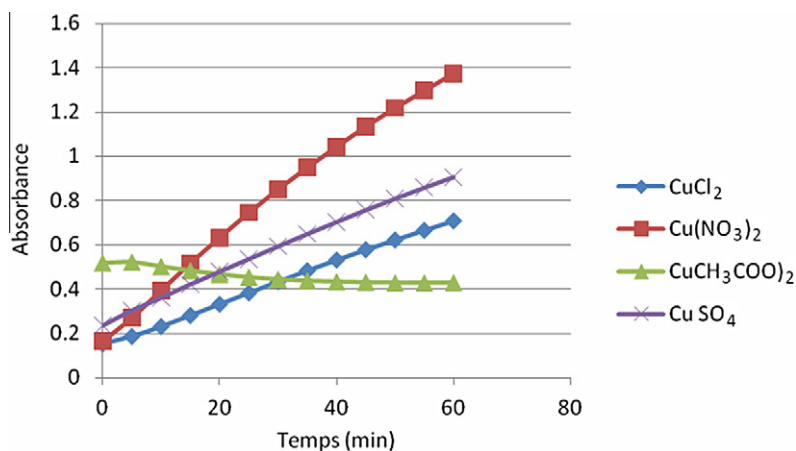


Figure 3 Oxidation of catechol by complexes of ligand L3.

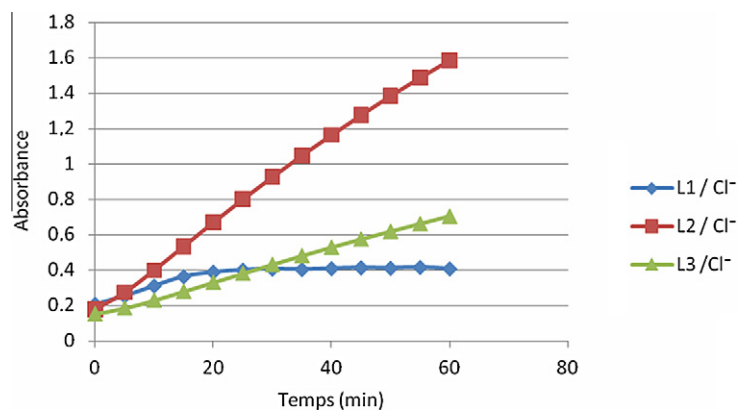


Figure 4 Oxidation of the catechol in presence of Cu(Cl)_2 .

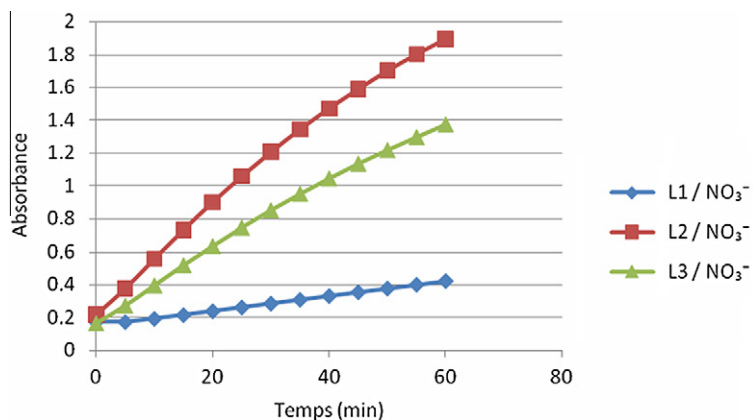


Figure 5 Oxidation of the catechol in presence of $\text{Cu(NO}_3)_2$.

at 65 °C for 4 h. The residual reactions were extracted with dichloromethane and washed with water. The organic solu-

tions were concentrated under reduced pressure to give the yellow oils.

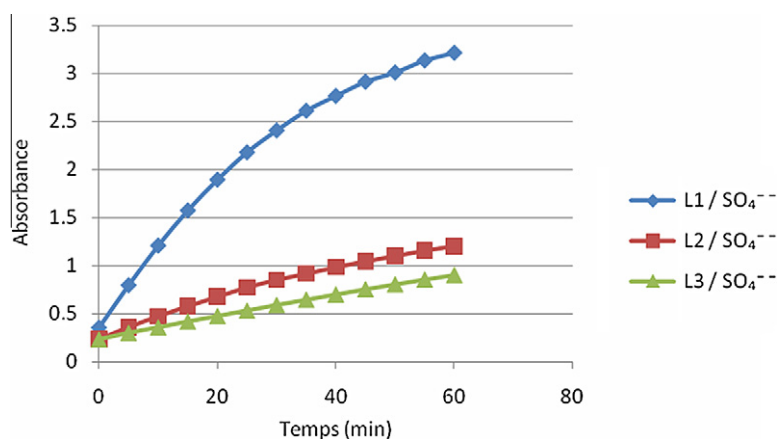


Figure 6 Oxidation of the catechol in presence of CuSO_4 .

Table 2 Oxidation rates ($\mu\text{mol L}^{-1} \text{min}^{-1}$) of catechol **L1**–**L3**.

Ligands	Copper salt ($2 \times 10^{-3} \text{ M}$)	V ($\text{mol L}^{-1} \text{min}^{-1}$)
L1	CuCl_2	1.80×10^{-6}
	$\text{Cu}(\text{NO}_3)_2$	2.77×10^{-6}
	$\text{Cu}(\text{CH}_3\text{COO})_2$	14.44×10^{-6}
	CuSO_4	28.99×10^{-6}
L2	CuCl_2	15.03×10^{-6}
	$\text{Cu}(\text{NO}_3)_2$	17.70×10^{-6}
	$\text{Cu}(\text{CH}_3\text{COO})_2$	—
	CuSO_4	9.9×10^{-6}
L3	CuCl_2	5.96×10^{-6}
	$\text{Cu}(\text{NO}_3)_2$	12.75×10^{-6}
	$\text{Cu}(\text{CH}_3\text{COO})_2$	—
	CuSO_4	7.01×10^{-6}

4.3. Characteristic data of new compounds

4.3.1. 3-(Bis(3,5-dimethyl-1H-pyrazol-1-yl)methyl)amino)propane-1-ol: **L1**

Yield: 80%; ^1H NMR (CDCl_3) δ ppm: 5.78 (s, 2 H, PzH); 4.90 (s, 2 H, $\text{N-CH}_2\text{-N}$); 4.36 (s, 2 H, $\text{N-CH}_2\text{-N}$); 3.81 (t, 2 H, $\text{CH}_2\text{-CH}_2\text{-OH}$, $J = 5.37 \text{ Hz}$); 2.96 (t, 2 H, $\text{CH}_2\text{-CH}_2\text{-N}$, $J = 5.63 \text{ Hz}$); 2.23 (s, 6 H, 2 CH_3); 2.23 (s, 6 H, 2 CH_3); 1.65 (m, 2 H, $\text{-CH}_2\text{-CH}_2\text{-CH}_2\text{-}$, $J = 5.46 \text{ Hz}$); ^{13}C NMR (CDCl_3) δ ppm: 147.68 ($\text{C}_{\text{Pz}}\text{-CH}_3$); 139.71 ($\text{CH}_3\text{-C}_{\text{Pz}}$); 105.75 ($\text{C}_{\text{Pz}}\text{H}$); 67.79 ($\text{N-CH}_2\text{-N}$); 64.89 ($\text{CH}_2\text{-OH}$); 47.21 ($\text{CH}_2\text{-N}$); 22.61 ($\text{CH}_2\text{-CH}_2\text{-CH}_2$); 12.09 (CH_3Pz); 0.97 (CH_3Pz); MS (ES) m/z : 291.73 (65%); 195.93 (100); 180.93 (5%); 97.20 (45%).

4.3.2. 1-[(3-Hydroxypropyl)[3-(methoxycarbonyl)-5-methyl-1H-pyrazol-1-yl]methylamino)methyl]-5-methyl-1H-pyrazole-3-carboxylate de méthyle: **L2**

Yield: 78%; ^1H NMR (CDCl_3) δ ppm: 6.56 (s, 2 H, PzH); 5.29 (s, 2 H, $\text{N-CH}_2\text{-N}$); 5.08 (s, 2 H, $\text{N-CH}_2\text{-N}$); 3.89 (s, 6 H, OCH_3); 3.82 (t, 2 H, $\text{CH}_2\text{-CH}_2\text{-OH}$, $J = 5.37 \text{ Hz}$); 2.97 (t, 2 H, $\text{CH}_2\text{-CH}_2\text{-N}$, $J = 5.62 \text{ Hz}$); 2.35 (s, 6 H, 2 CH_3); 1.65 (m, 2 H, $\text{-CH}_2\text{-CH}_2\text{-CH}_2\text{-}$, $J = 5.55 \text{ Hz}$); ^{13}C NMR (CDCl_3) δ ppm: 163.09 (CO); 142.19 ($\text{C}_{\text{Pz}}\text{=N}$); 140.70 ($\text{C}_{\text{Pz}}\text{=C}$); 108.96 ($\text{C}_{\text{Pz}}\text{H}$); 67.79 ($\text{N-CH}_2\text{-N}$); 66.93 ($\text{CH}_2\text{-OH}$); 51.90 ($\text{CH}_3\text{-O}$); 47.10 ($\text{CH}_2\text{-N}$); 22.63 ($\text{CH}_2\text{-CH}_2\text{-CH}_2$); 11.20 (CH_3Pz); MS

(ES) m/z (%): 379.64 (10); 337.60 (25); 294.80 (50); 280.87 (100); 239.93 (25); 184.20 (30); 141.07 (45); 100.07 (20).

4.3.3. 1-[(3-Hydroxypropyl)[3-(ethoxycarbonyl)-5-methyl-1H-pyrazol-1-yl]methylamino)methyl]-5-methyl-1H-pyrazole-3-carboxylate d'ethyle: **L3**

Yield: 75%; ^1H NMR (CDCl_3) δ ppm: 6.55 (s, 2 H, $\text{C}_{\text{Pz}}\text{H}$); 5.29 (s, 2 H, $\text{N-CH}_2\text{-N}$); 5.06 (s, 2 H, $\text{N-CH}_2\text{-N}$); 4.30 (q, 4 H, $\text{O-CH}_2\text{-CH}_3$, $J = 7.1 \text{ Hz}$); 3.62 (t, 2 H, $\text{CH}_2\text{-CH}_2\text{-OH}$, $J = 5.03 \text{ Hz}$); 2.99 (t, 2 H, $\text{CH}_2\text{-CH}_2\text{-N}$, $J = 5.00 \text{ Hz}$); 2.27 (s, 6 H, 2 CH_3); 1.33 (t, 6 H, $\text{-CH}_2\text{-CH}_3$, $J = 7.1 \text{ Hz}$); 1.65 (m, 2 H, $\text{-CH}_2\text{-CH}_2\text{-CH}_2\text{-}$, $J = 5.55 \text{ Hz}$); ^{13}C NMR (CDCl_3) δ ppm: 162.67 (CO); 142.51 ($\text{C}_{\text{Pz}}\text{=N}$); 140.59 ($\text{C}_{\text{Pz}}\text{=C}$); 108.85 ($\text{C}_{\text{Pz}}\text{H}$); 67.76 ($\text{N-CH}_2\text{-N}$); 66.82 ($\text{CH}_2\text{-OH}$); 60.78 ($\text{CH}_2\text{-O}$); 47.10 ($\text{CH}_2\text{-N}$); 22.62 ($\text{CH}_2\text{-CH}_2\text{-CH}_2$); 14.24 ($\text{-CH}_2\text{-CH}_3$); 11.16 (CH_3Pz); MS (ES) m/z (%): 407.47 (5); 308.8 (100); 294.93 (15); 250.93 (15); 155.07 (15); 97.20 (5).

4.4. Catecholase activity measurements

Kinetic measurements were made spectrophotometrically on a UV-Visible spectrophotometer (In the COSTE: Centre de l'Oriental des Sciences et Technologies de l'Eau), following the appearance of *o*-quinone over time at 25 °C (390 nm absorbance maximum, $\epsilon = 1600 \text{ M}^{-1} \text{cm}^{-1}$ in methanol). The metal complex (prepared *in situ* from copper(II) salt and the ligand, 0.3 mL of 10^{-3} M methanol solution) and a 2 mL solution (10^{-1} M methanol solution) of catechol were added together in the spectrophotometric cell.

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References

- Albada, H.B., Soulimani, F., Weckhuysen, B.M., Liskamp, R.M.J., 2007. Scaffolded amino acids as a close structural mimic of type-3 copper binding sites. *Chem. Commun.* 46, 4895.

- Bouabdallah, I., Touzani, R., Zidane, I., Ramdani, A., 2007. Synthesis of new tripodal ligand: N,N-bis[(1,5-dimethyl-3-yl)methyl] benzylamine. Catecholase activity of two series of tripodal ligands with some copper(II) salts. *Catal. Commun.* 8, 707.
- Boussalah, N., Touzani, R., Bouabdallah, I., El Kadiri, S., Ghalem, S., 2009a. Oxidation catalytic properties of new amino acid based on pyrazole tripodal ligands. *Int. J. Aca. Res.* 2, 137.
- Boussalah, N., Touzani, R., Bouabdallah, I., El Kadiri, S., Ghalem, S., 2009b. Synthesis, structure and catalytic properties of tripodal amino-acid derivatized pyrazole-based ligands. *J. Mol. Cat. A Chem.* 306, 113.
- Chen, P., Solomon, E.I., 2004. Oxygen activation by the noncoupled binuclear copper site in peptidylglycine α -hydroxylating monooxygenase. Reaction mechanism and role of the noncoupled nature of the active site. *J. Am. Chem. Soc.* 126, 4991.
- Decker, H., Tuczek, F., 2000. Tyrosinase/catecholoxidase activity of hemocyanins: structural basis and molecular mechanism. *Trends Biochem. Sci.* 25, 392.
- Decker, H., Dillinger, R., Tuczek, F., 2000. How does tyrosinase work? Recent insights from model chemistry and structural biology. *Angew. Chem. Int. Ed.* 39, 1591.
- Dvoretzky, I., Richter, G.H., 1950. Formaldehyde condensation in the pyrazole series. *J. Org. Chem.* 15, 1285.
- El Kodadi, M., Malek, F., Touzani, R., Ramdani, A., 2008. Synthesis of new tripodal ligand 5-(bis(3,5-dimethyl-1H-pyrazol-1-ylmethyl)amino)pentan-1-ol, catecholase activities studies of three functional tripodal pyrazolyl N-donor ligands, with different copper(II) salts. *Catal. Commun.* 9, 966.
- Gerdemann, C., Eicken, C., Krebs, B., 2002. The crystal structure of catechol oxidase: new insight into the function of type-3 copper proteins. *Acc. Chem. Res.* 35, 183.
- Holm, R.H., Kennepohl, P., Solomon, E.I., 1996. Structural and functional aspects of metal sites in biology. *Chem. Rev.* 96, 2239.
- Kitajima, N., Moro-oka, Y., 1994. Copper-dioxygen complexes. Inorganic and bioinorganic perspectives. *Chem. Rev.* 94, 737.
- Koval, I.A., Selmececi, K., Belle, C., Philouze, C., Saint-Aman, E., Gautier-Luneau, I., Schuitema, A.M., Van Vliet, M., Gamez, P., Roubeau, O., Lüken, M., Krebs, B., Lutz, M., Spek, A.L., Pierre, J.-L., Reedijk, J., 2006. Catecholase activity of a copper(II) complex with a macrocyclic ligand: unraveling catalytic mechanisms. *Chem. – A Eur. J.* 12, 6138.
- Malachowski, M.R., Dorsey, B., Sackett, J.G., Kelly, R.S., Ferko, A.L., Hardin, R.N., 1996. Effect of ligand donors on the catalytic properties of metal complexes. Copper(II) complexes as catalysts for the oxidation of 3,5-di-tert-butylcatechol. *Inorg. Chim. Acta* 249, 85.
- Rompel, A., Fischer, H., Meiwes, D., Büldt-Karentzopoulos, K., Dillinger, R., Tuczek, F., Witzel, H., Krebs, B., 1999. Purification and spectroscopic studies on catechol oxidases from *Lycopus europaeus* and *Populus nigra*: evidence for a dinuclear copper center of type 3 and spectroscopic similarities to tyrosinase and hemocyanin. *J. Biol. Inorg. Chem.* 4, 56.
- Touzani, R., Ramdani, A., El Kadiri, S., Gourand, F., 1999. 1-Hydroxymethyl-3-(ethoxycarbonyl)-5-methylpyrazole. *Molecules* 4, M116.
- Touzani, R., Ramdani, A., El Kadiri, S., 2000. 1-Hydroxymethyl-3-methoxycarbonyl-5-methylpyrazole. *Molecules* 5, M139.
- Touzani, R., Ramdani, A., Ben-Hadda, T., El Kadiri, S., Maury, O., Le Bozec, H., Dixneuf, P.H., 2001. Efficient synthesis of new nitrogen donor containing tripods under microwave irradiation and without solvent. *Synth. Commun.* 31, 1315.
- Van Gelder, C.W.G., Flurkey, W.H., Wichers, H.J., 1997. Sequence and structural features of plant and fungal tyrosinases. *Phytochemistry* 45, 1309.